

From Lots of Pictures to Lots of Numbers to Lots of Answers:

Advances in quantitative high- content imaging

Dr. Erin Weisbart

Senior Computational Scientist, Lead Image Analyst

Cimini Lab, Imaging Platform

Broad Institute of MIT and Harvard





Overview

Why images?

**Advances in software
data science
machine learning**

**Importance of public data
open-source software
cross-discipline collaboration**



Why images?

Biological images are informative

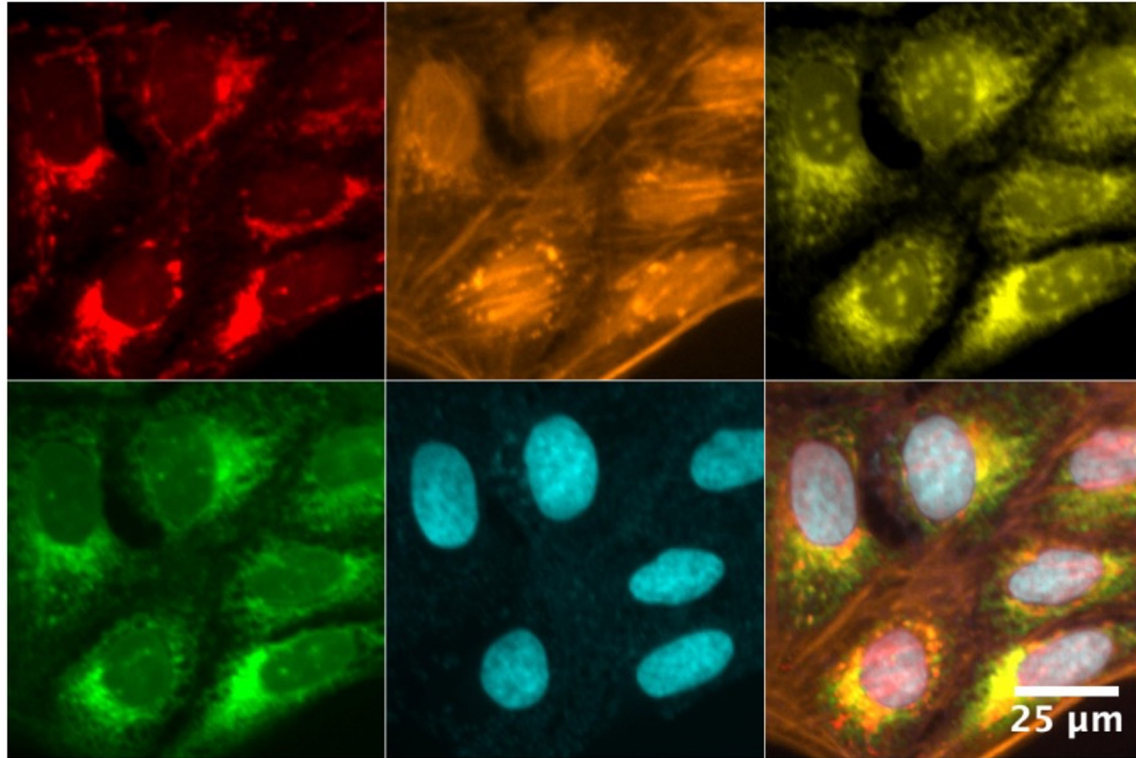
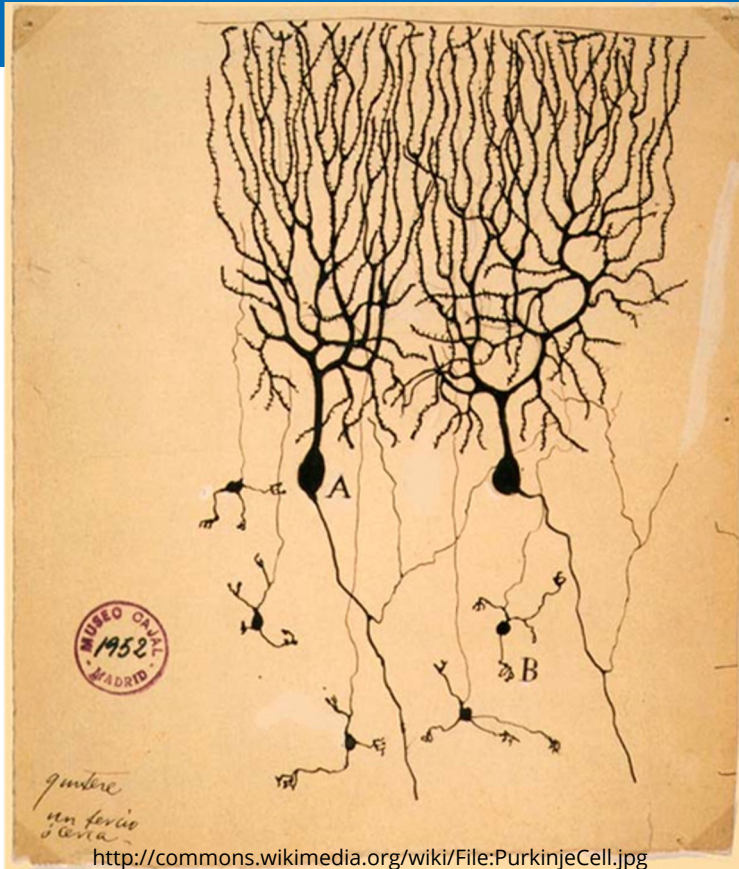


Image data can be:

- Single cell resolution
- Interaction between molecules
- Live cell data
- Changes in expression level **and** behavior
- ...
- Quantitative

Microscopy's history is mostly qualitative



Drawing of Purkinje cells (A) and granule cells (B) from pigeon cerebellum, 1899. Instituto Santiago Ramón y Cajal, Madrid, Spain

My history of image quantitation

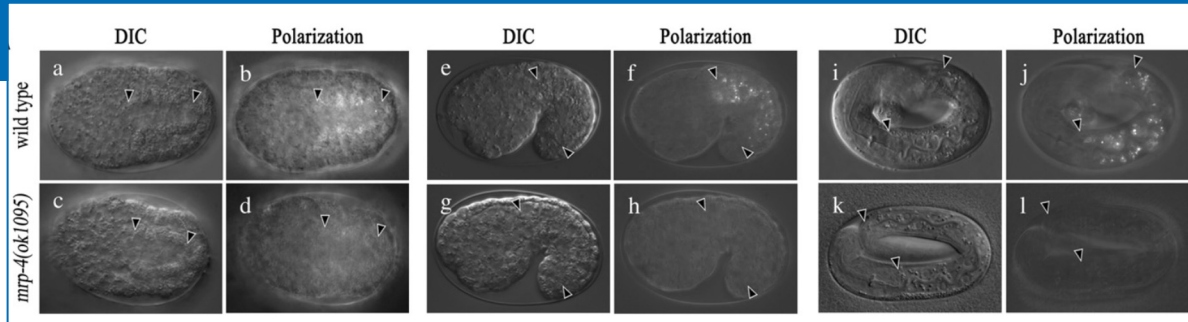


TABLE 1

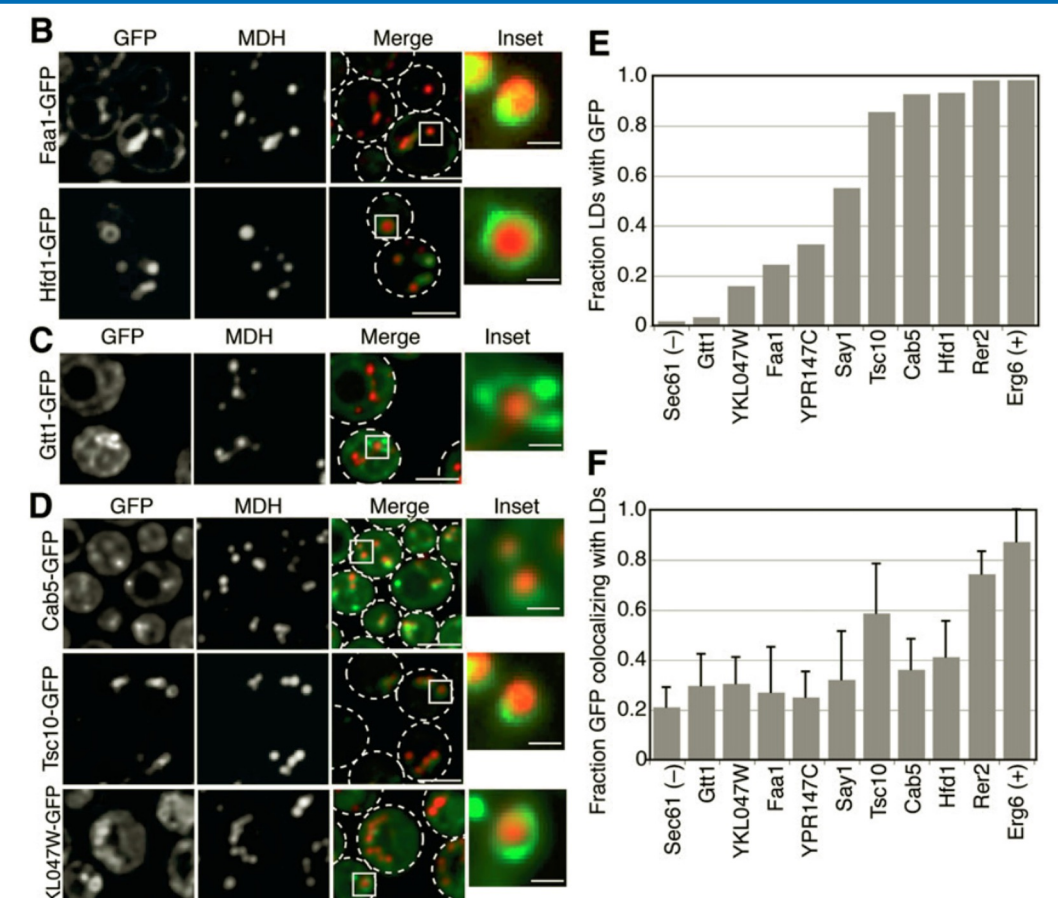
Analysis of birefringent gut granules

Genotype	% of embryos lacking birefringent material in intestinal cells (<i>n</i>)	% of larvae lacking birefringent material in intestinal cells (<i>n</i>)
Wild type ^a	0 (>100)	0 (>100)
ABC transporters		
<i>mrp-4(cd8)</i>	93 (99)	20 (46)
<i>mrp-4(ok1095)</i>	96 (164)	12 (182)
<i>mrp-4(RNAi)</i>	93 (390)	14 (227)
<i>wht-2(RNAi)</i>	52 (673)	5 (137)
Double mutants		
<i>mrp-4(cd8); whe-2(RNAi)</i>	97 (107)	81 (89)
<i>mrp-4(ok1095); whe-2(RNAi)</i>	100 (109)	89 (61)
Mosaic RNAi		
<i>rrf-1(pk1471)^a</i>	0 (96)	0 (47)
<i>rrf-1(pk1471); mrp-4(RNAi)</i>	53 (45)	36 (22)
<i>rrf-1(pk1471); whe-2(RNAi)</i>	3 (119)	0 (26)
<i>rsd-2(pk3307)^a</i>	0 (84)	0 (45)
<i>rsd-2(pk3307); mrp-4(RNAi)</i>	0 (63)	0 (40)

Currie, E., King, B., Lawrenson, A. L., Schroeder, L. K., Kershner, A. M., & Hermann, G. J. (2007). Role of the *Caenorhabditis elegans* multidrug resistance gene, *mrp-4*, in gut granule differentiation. *Genetics*, 177(3), 1569–1582. <https://doi.org/10.1534/genetics.107.080689>

My history of image quantitation

Currie, E., Guo, X., Christiano, R., Chitraju, C., Kory, N., Harrison, K., Haas, J., Walther, T. C., & Farese, R. V. (2014). High confidence proteomic analysis of yeast LDs identifies additional droplet proteins and reveals connections to dolichol synthesis and sterol acetylation [S]. *Journal of Lipid Research*, 55(7), 1465–1477. <https://doi.org/10.1194/jlr.M050229>



Imaging Platform at Broad Institute



Anne E. Carpenter
Senior Director of the Imaging Platform,
Institute Scientist



Shantanu Singh
Senior Group Leader



Beth Cimini
Associate Director for Bioimage Analysis

Software to turn Images → Numbers → Answers



CellProfiler™
cell image analysis software



CellProfiler Analyst™
data exploration software



**Distributed
Something™**



PyCytominer

PIXIMI™



How has software
and data science
changed imaging?

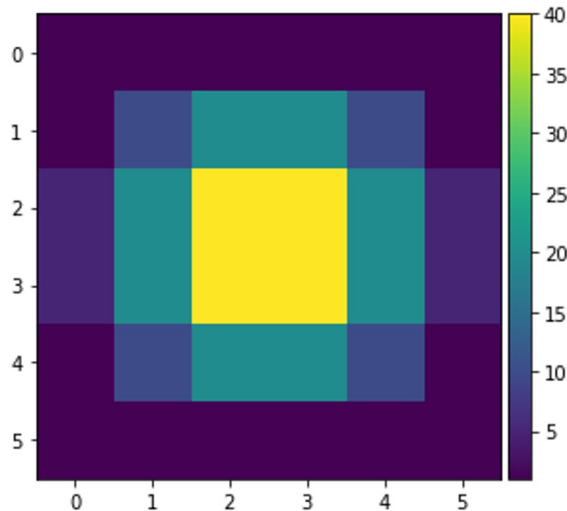


Digital images are arrays

```
skimage.io.imshow(a)
```

```
/usr/local/lib/python3.8/site-packages/skimage  
e; displaying image with stretched contrast.  
  lo, hi, cmap = _get_display_range(image)
```

```
<matplotlib.image.AxesImage at 0x125202070>
```



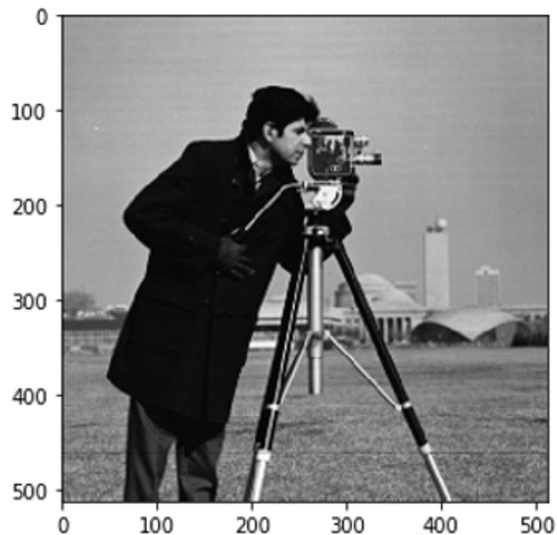
```
a
```

```
array([[ 1,  1,  1,  1,  1,  1],  
       [ 1, 10, 20, 20, 10,  1],  
       [ 5, 20, 40, 40, 20,  5],  
       [ 5, 20, 40, 40, 20,  5],  
       [ 1, 10, 20, 20, 10,  1],  
       [ 1,  1,  1,  1,  1,  1]])
```

Digital images are arrays

```
skimage.io.imshow(b)
```

```
<matplotlib.image.AxesImage at 0x12552cca0>
```



b

```
array([[156, 157, 160, ..., 152, 152, 152],  
       [156, 157, 159, ..., 152, 152, 152],  
       [158, 157, 156, ..., 152, 152, 152],  
       ...,  
       [121, 123, 126, ..., 121, 113, 111],  
       [121, 123, 126, ..., 121, 113, 111],  
       [121, 123, 126, ..., 121, 113, 111]], dtype=uint8)
```



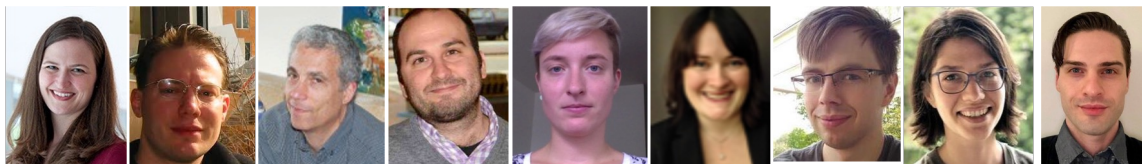
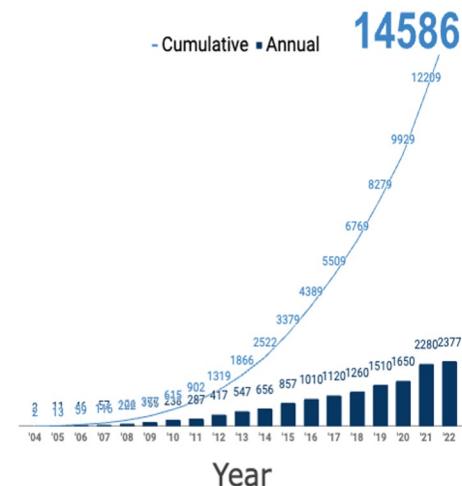
CellProfiler™
cell image analysis software



Open-source software for image analysis

- Free and open-source; Windows, Mac, Linux
- Ranked **most flexible** and **usable** in independent analysis (*Wiesmann et al.*)
- Cited in **2,000+** papers per year
- Used in **7/10** top pharma companies
- In the **Top 10** most popular papers in Genome Biology

Publications citing CellProfiler



Anne
Carpenter

Ray
Jones

Lee
Kamentsky

Allen
Goodman

Claire
McQuin

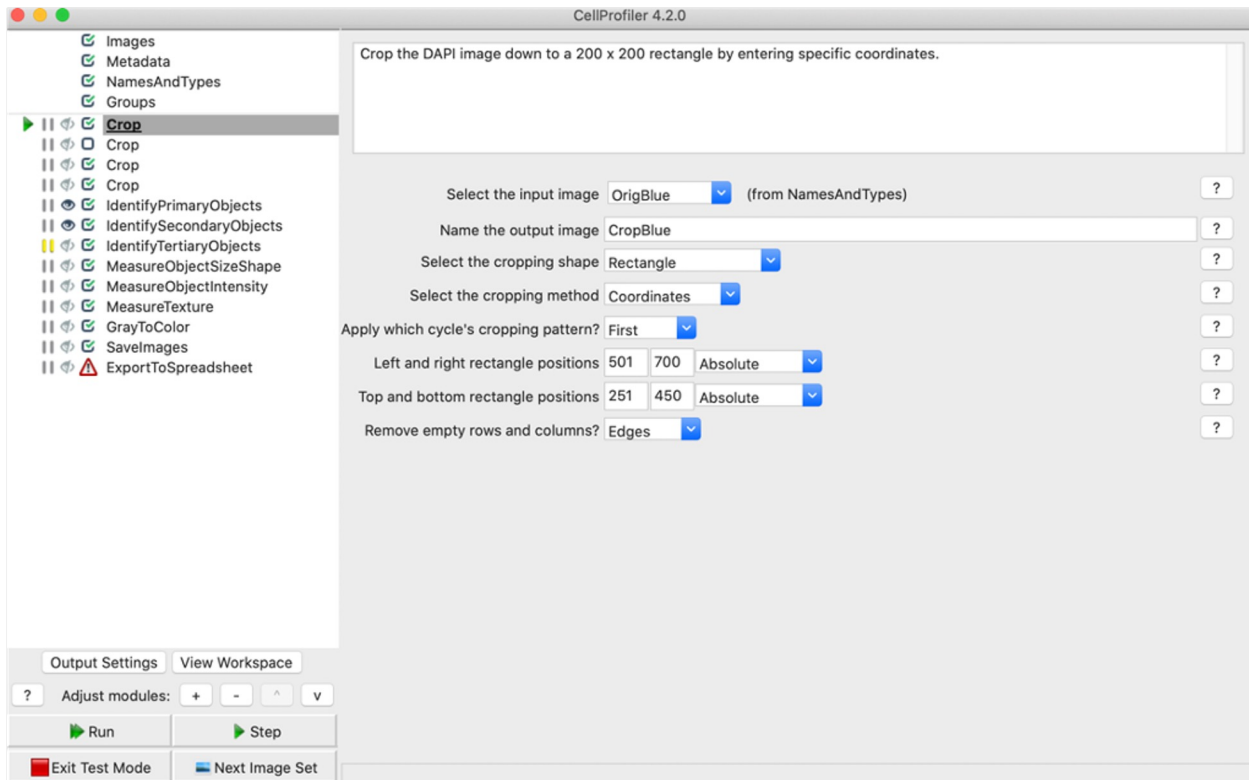
Beth
Cimini

David
Stirling

Alice
Lucas

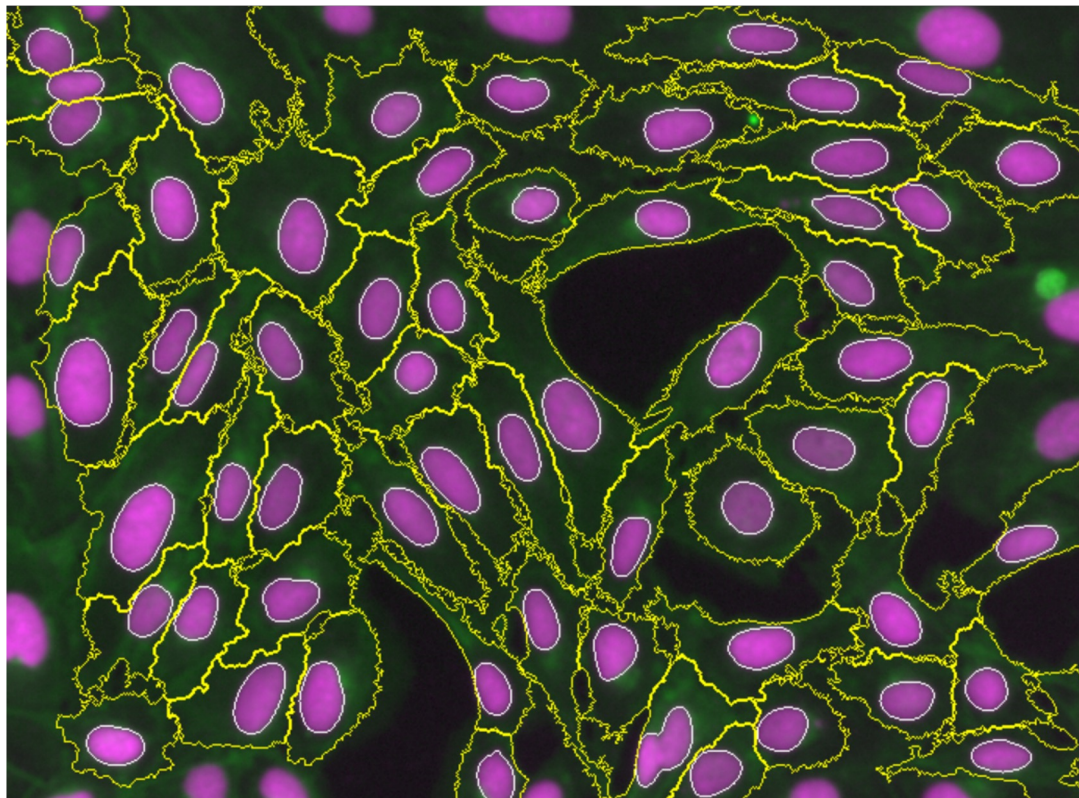
Nodar
Gogoberidze

CellProfiler Interface



- 94 “modules”
- 930 settings
- Total library is 121K lines of code, 37K lines of documentation (23% documentation)

Identify Objects





Measure object ... intensity

- Integrated intensity: Sum of the pixel intensities within an object
- Mean, median, standard deviation intensities
- Maximal and minimal pixel intensities
- Lower/Upper quartile of the intensity
- Object intensities may be measured from any channel, not just the channel used to identify the object

Measure object ... intensity distribution

Calculate intensity Magnitudes and phase

Maximum zernike moment 9

Select an image to measure image_input (from NamesAndTypes)
Add another image

Select objects to measure Cell (from IdentifySecondaryObjects)

Object to use as center? These objects
Add another object

Scale the bins? Yes No

Number of bins 4

Scale the bins? Yes No

Number of bins 4

Maximum radius 35

Magnitude:
Amount of intensity within each ring

Phase (Zernike moment):
Distribution of intensity within each ring

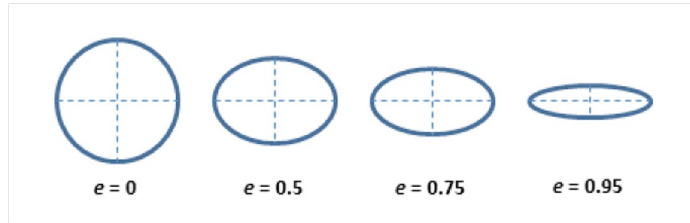
E.g. one side of the cell is brighter

Within each fraction/ring:
- *FracAtD*: total intensity
- *MeanFrac*: mean intensity
- *RadialCV*: divide the ring into 8 slices, measure the coefficient of variation

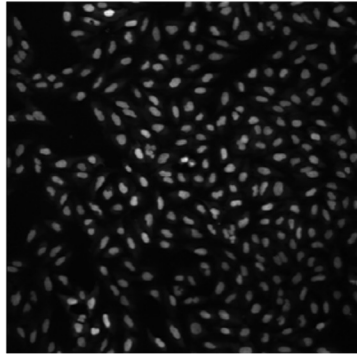
Measure object ... size and shape



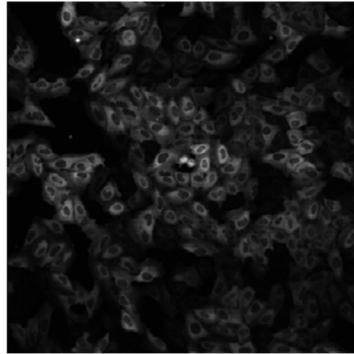
- Area
- Perimeter
- Eccentricity (circle = 1, line = 0)
- MajorAxisLength
- MinorAxisLength
- Orientation
- Solidity



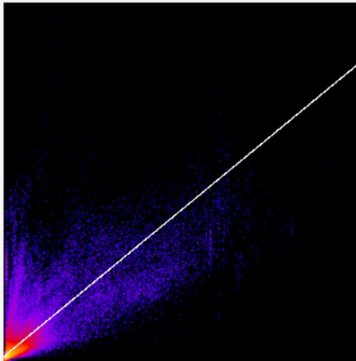
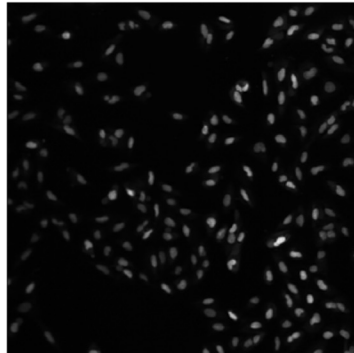
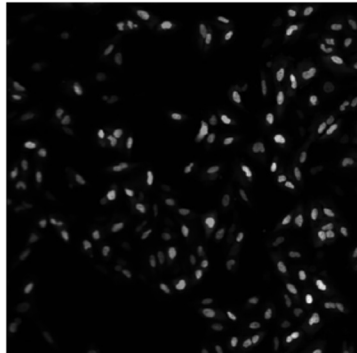
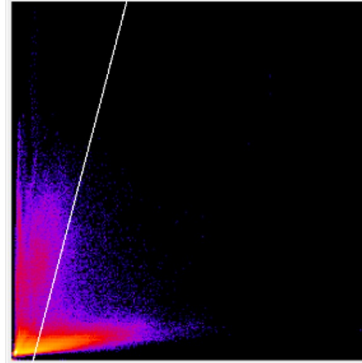
Measure object ... colocalization



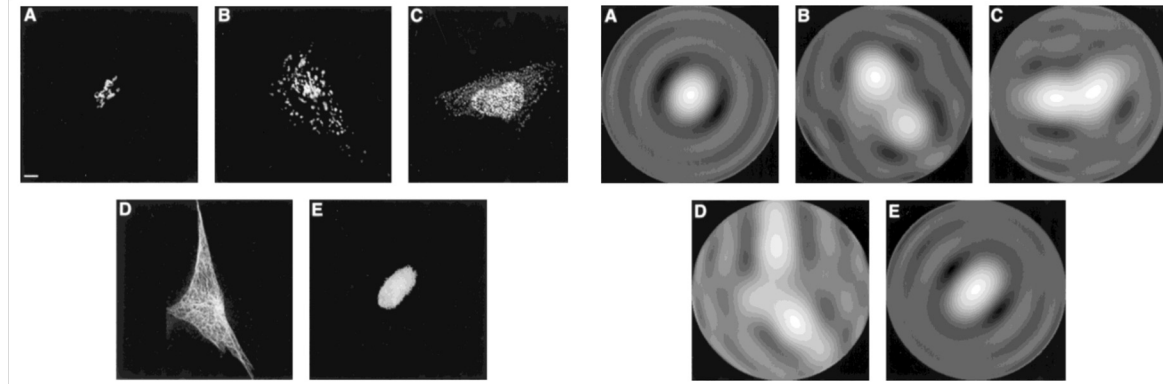
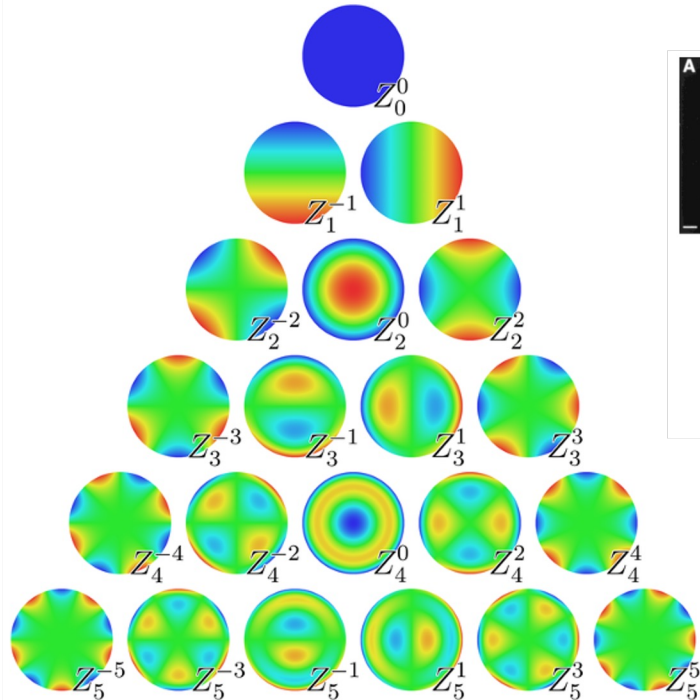
DAPI



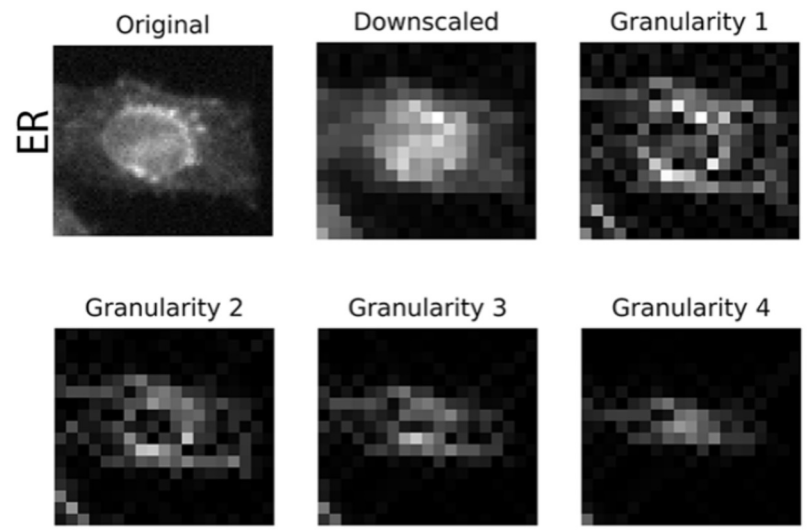
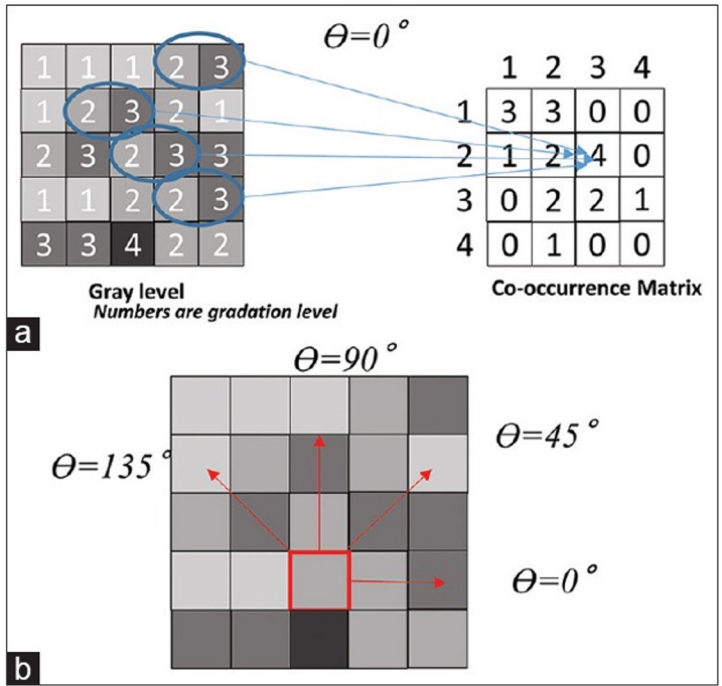
GFP



Measure object ... Zernike polynomials



Measure object ... texture, granularity



Most high-content screens are not data rich

60-80% of “high-content” studies use only 1 or 2 cellular features

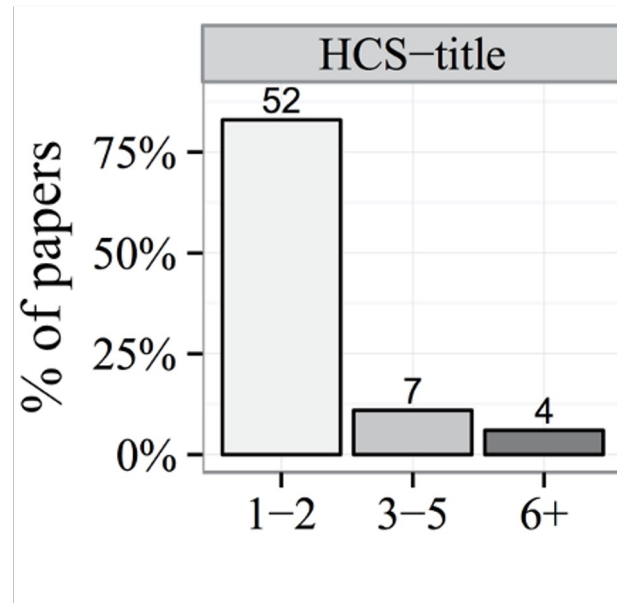
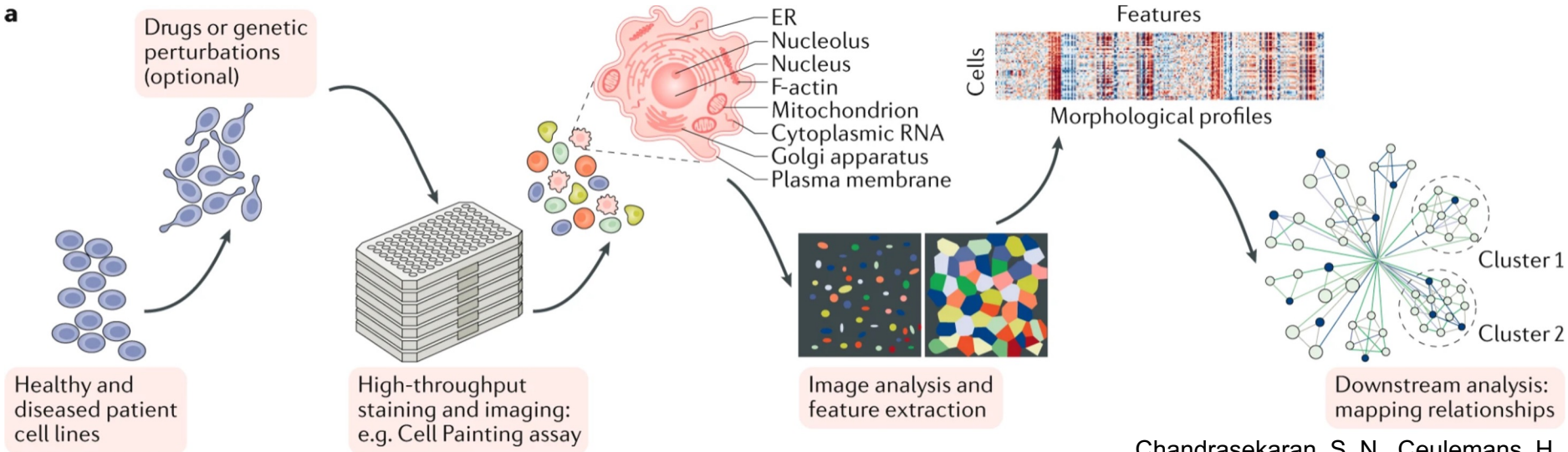


Image-based (Morphological) Profiling

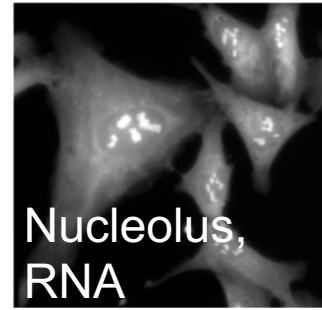
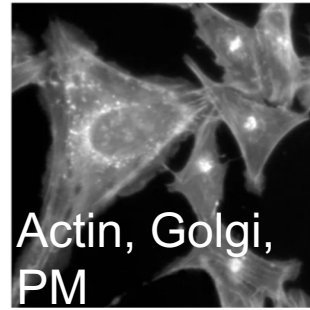
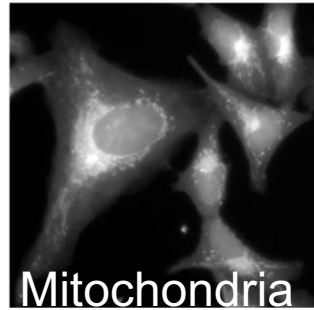
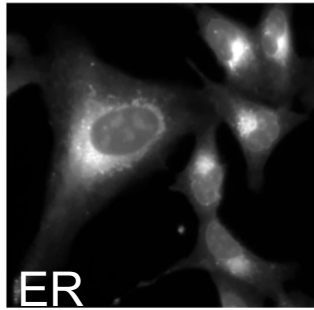
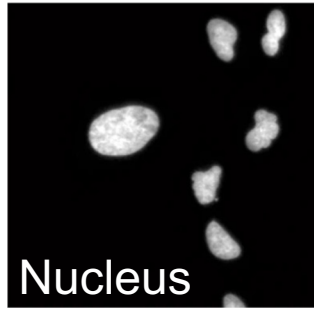
a



Chandrasekaran, S. N., Ceulemans, H., Boyd, J. D., & Carpenter, A. E. (2021). Image-based profiling for drug discovery: due for a machine-learning upgrade? *Nature Reviews. Drug Discovery*, 20(2), 145–159. <https://doi.org/10.1038/s41573-020-00117-w>

Image-based Profiling with Cell Painting

Cell Painting Assay



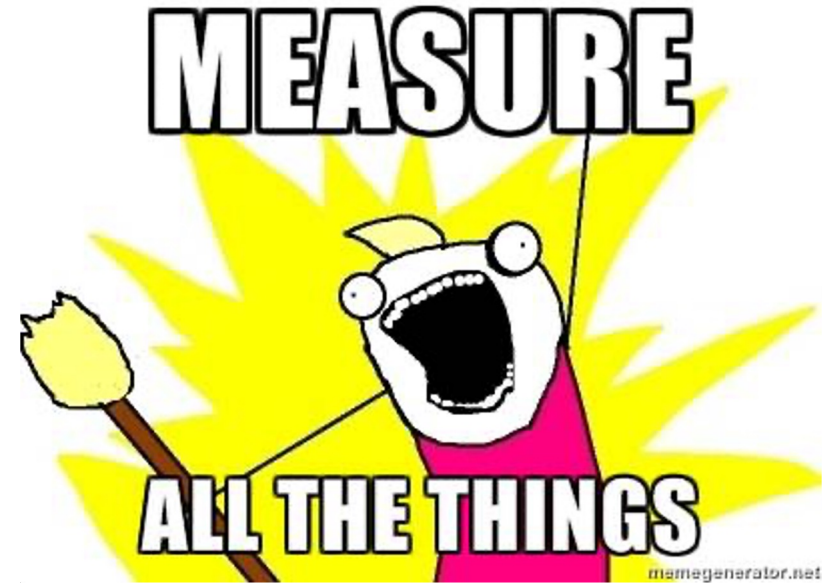
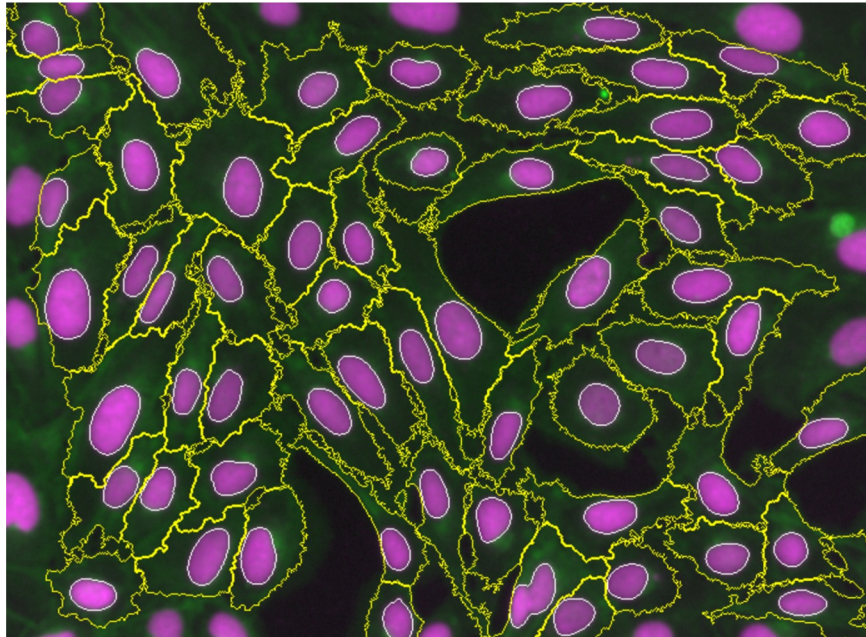
6 stains, 5 channels, 8 organelles/compartments

½ to ¼ cost of mRNA profiling (L1000)

Cimini, B. A., Chandrasekaran, S. N., Kost-Alimova, M., Miller, L., Goodale, A., Fritchman, B., Byrne, P., Garg, S., Jamali, N., Logan, D. J., Concannon, J. B., Lardeau, C.-H., Mouchet, E., Singh, S., Shafqat Abbasi, H., Aspesi, P., Jr, Boyd, J. D., Gilbert, T., Gnutt, D., ... Carpenter, A. E. (2023). Optimizing the Cell Painting assay for image-based profiling. *Nature Protocols*.
<https://doi.org/10.1038/s41596-023-00840-9>

Image-based Profiling with Cell Painting

Identify nuclei, cells, cytoplasm

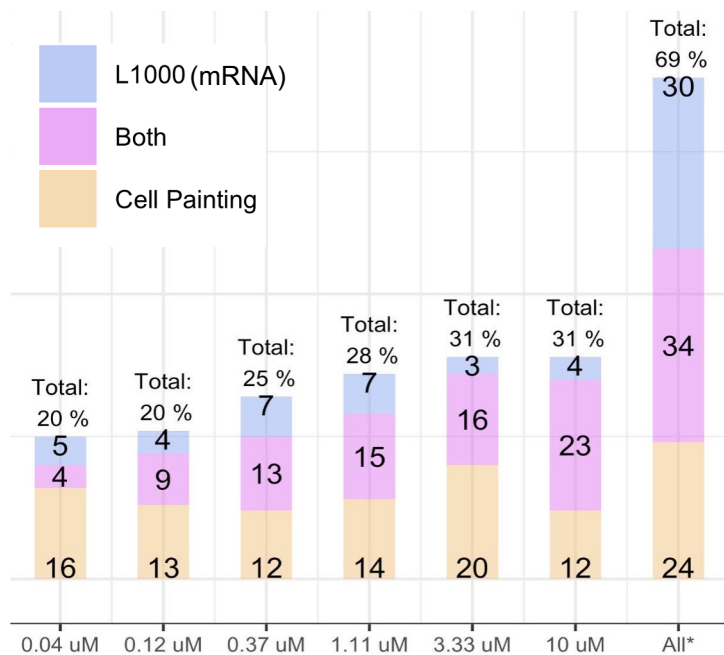


Cell Painting used for:

- **predicting small molecule activity**
- **characterizing gene alleles within cells**
- **predicting cellular drug resistance**
- **distinguishing between disease states**
- **predicting outcomes of other assays**
- **...**

Cell Painting complements mRNA profiling

Unsupervised analysis: number (and %) of MOA classes whose compounds are self-similar



Way, G. P., Natoli, T., Adeboye, A., Litichevskiy, L., Yang, A., Lu, X., Caicedo, J. C., Cimini, B. A., Karhohs, K., Logan, D. J., Rohban, M. H., Kost-Alimova, M., Hartland, K., Bornholdt, M., Chandrasekaran, S. N., Haghighi, M., Weisbart, E., Singh, S., Subramanian, A., & Carpenter, A. E. (2022). Morphology and gene expression profiling provide complementary information for mapping cell state. *Cell Systems*, 13(11), 911–923.e9. <https://doi.org/10.1016/j.cels.2022.10.001>

Cell Painting complements protein profiling



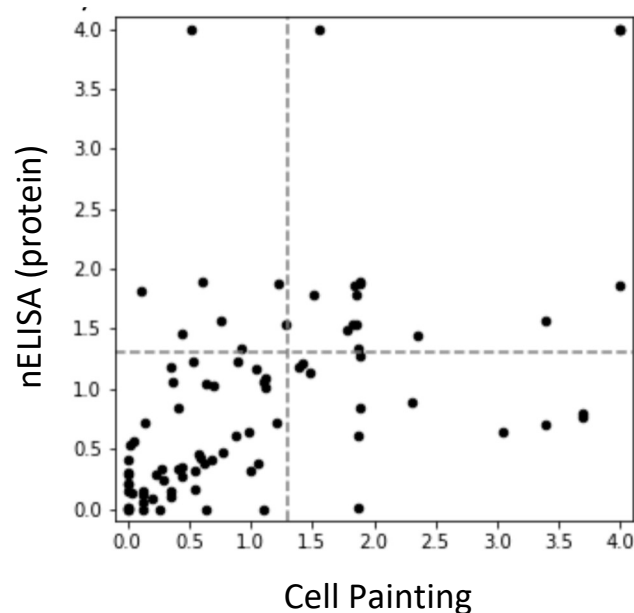
nELISA by Nomic bio

A next-generation, massively parallel ELISA

Current capabilities:

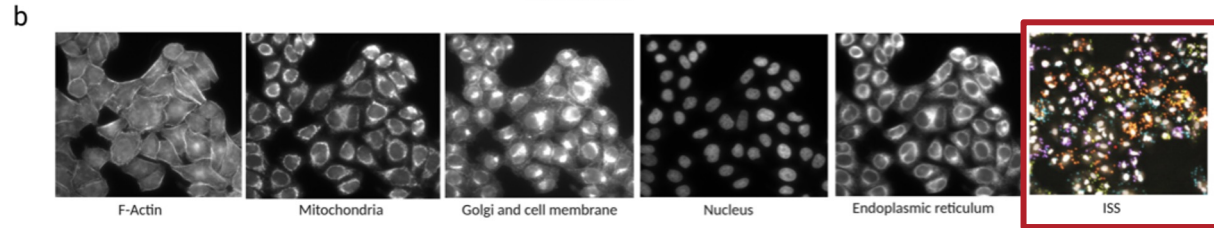
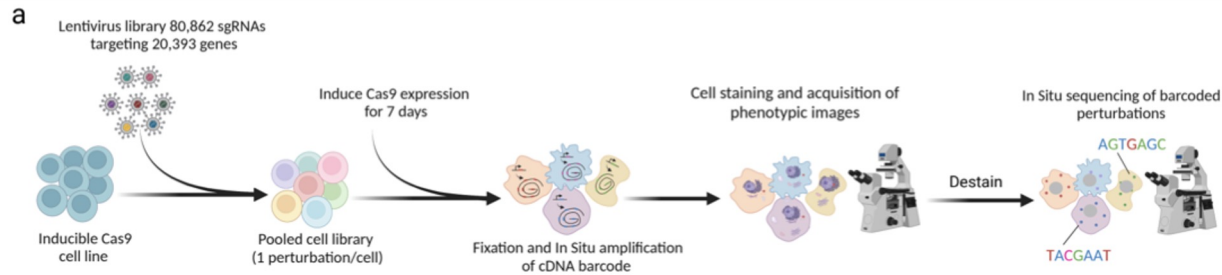
- 191-plex secretome panel

Predicting the mechanism of action for compounds

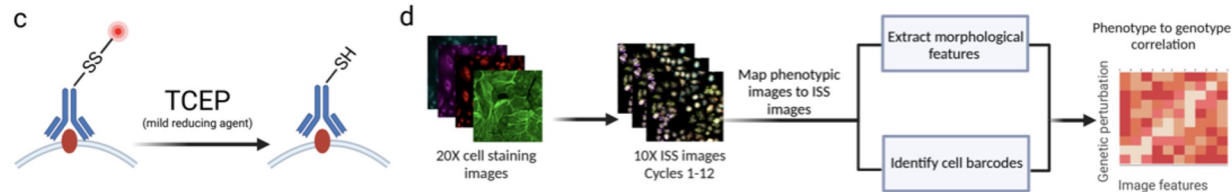


Dagher, M., Ongo, G., Robichaud, N., Kong, J., & Rho, W. (2023). nELISA: A high-throughput, high-plex platform enables quantitative profiling of the secretome. *bioRxiv*.
<https://www.biorxiv.org/content/10.1101/2023.04.17.535914.abstract>

Pooled Cell Painting (PERISCOPE)

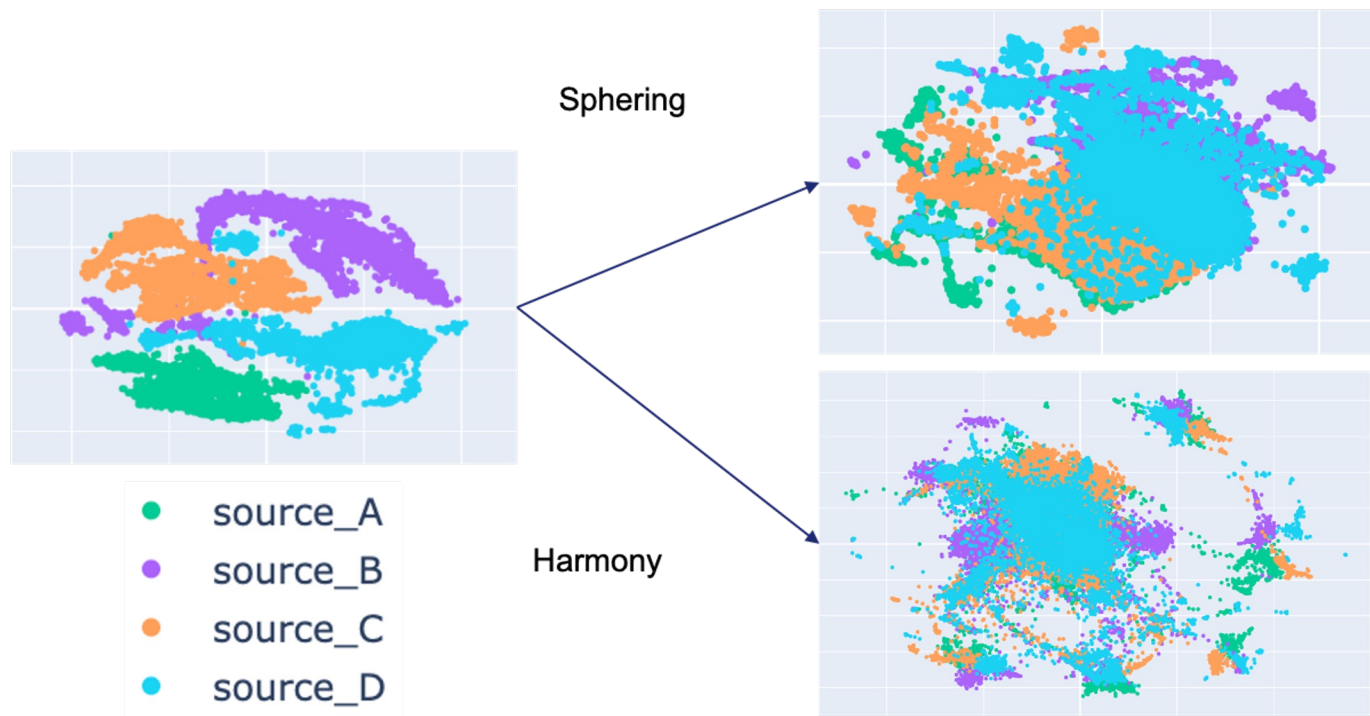


cost:
~\$0.001/cell




Ramezani, M.* , Bauman, J.* , Singh, A.* , **Weisbart, E.*** , Yong, J., Lozada, M., Way, G. P., Kavari, S. L., Diaz, C., Haghghi, M., Batista, T. M., Pérez-Schindler, J., Claussnitzer, M., Singh, S., Cimini, B. A., Blainey, P. C., Carpenter, A. E., Jan, C. H., & Neal, J. T. (2023). A genome-wide atlas of human cell morphology. In bioRxiv (p. 2023.08.06.552164). <https://doi.org/10.1101/2023.08.06.552164>


Bringing tools from other -omics into morphological profiling: Harmony



Arevalo, J., van Dijk, R., Carpenter, A. E., & Singh, S. (2023). Evaluating batch correction methods for image-based cell profiling. *bioRxiv : The Preprint Server for Biology*.
<https://doi.org/10.1101/2023.09.15.558001>



How can we bring
quantitative bioimaging
to everyone?



FAIR data principles

Findable

- (meta)data are assigned a globally unique and persistent identifier
- data are described with rich metadata
- metadata clearly and explicitly include the identifier of the data it describes
- (meta)data are registered or indexed in a searchable resource

Accessible

- (meta)data are retrievable by their identifier using a standardized communications protocol
- the protocol is open, free, and universally implementable
- the protocol allows for an authentication and authorization procedure, where necessary
- metadata are accessible, even when the data are no longer available

Interoperable

- (meta)data use a formal, accessible, shared, and broadly applicable language for knowledge representation
- (meta)data use vocabularies that follow FAIR principles
- (meta)data include qualified references to other (meta)data

Reusable

- meta(data) are richly described with a plurality of accurate and relevant attributes
- (meta)data are released with a clear and accessible data usage license
- (meta)data are associated with detailed provenance
- (meta)data meet domain-relevant community standards

Cell Painting Gallery on RODA

Registry of Open Data on AWS



Cell Painting Gallery

bioinformatics biology cancer cell biology cell imaging cell painting chemical biology computer vision csv deep learning fluorescence imaging genetic high-throughput imaging image processing image-based profiling imaging machine learning medicine microscopy organelle

Description

The Cell Painting Gallery is a collection of image datasets created using the [Cell Painting](#) assay. The images of cells are captured by microscopy imaging, and reveal the response of various labeled cell components to whatever treatments are tested, which can include genetic perturbations, chemicals or drugs, or different cell types. The datasets can be used for diverse applications in basic biology and pharmaceutical research, such as identifying disease-associated phenotypes, understanding disease mechanisms, and predicting a drug's activity, toxicity, or mechanism of action ([Chandrasekaran et al 2020](#)). This collection is maintained by the [Carpenter–Singh lab](#) and the [Cimini lab](#) at the [Broad Institute](#). A human-friendly listing of datasets, instructions for accessing them, and other documentation is at the [corresponding GitHub page](#) about the Gallery.

Update Frequency

Typically when an associated publication is posted on biorxiv

License

CC0 1.0 Universal (CC0 1.0) Public Domain Dedication, but please do cite the corresponding publication for each dataset, as listed [here](#).

Documentation

<https://github.com/broadinstitute/cellpainting-gallery>

Managed By:

Resources on AWS

Description

Cell Painting data, comprising fluorescence microscopy cell images (TIFF), extracted features (CSV), and associated metadata (CSV and TXT).

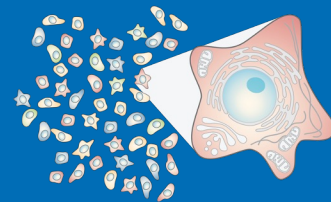
18 complete datasets

13 in-progress datasets

671.1 TB data

JUMP in Cell Painting Gallery

Joint Undertaking for Morphological Profiling

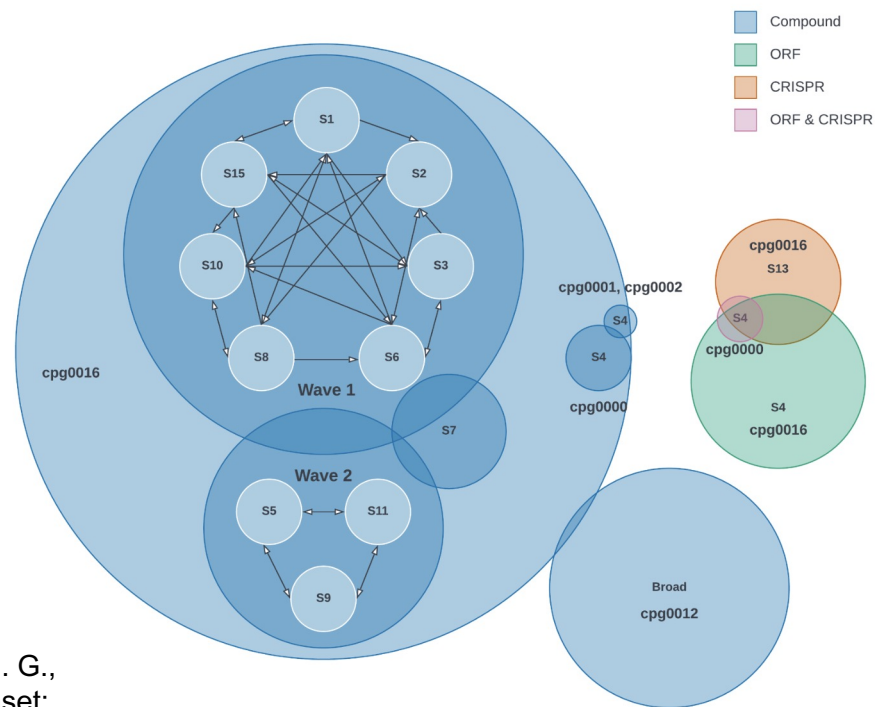


13,000 overexpressed genes

8,000 CRISPR knockdowns

116,000 (+ 30,000) small molecules

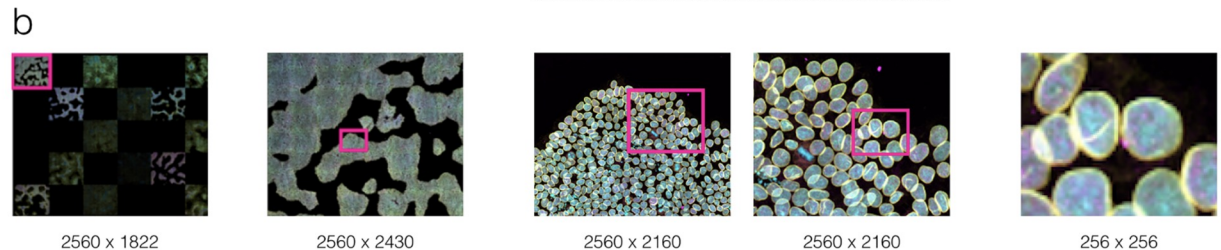
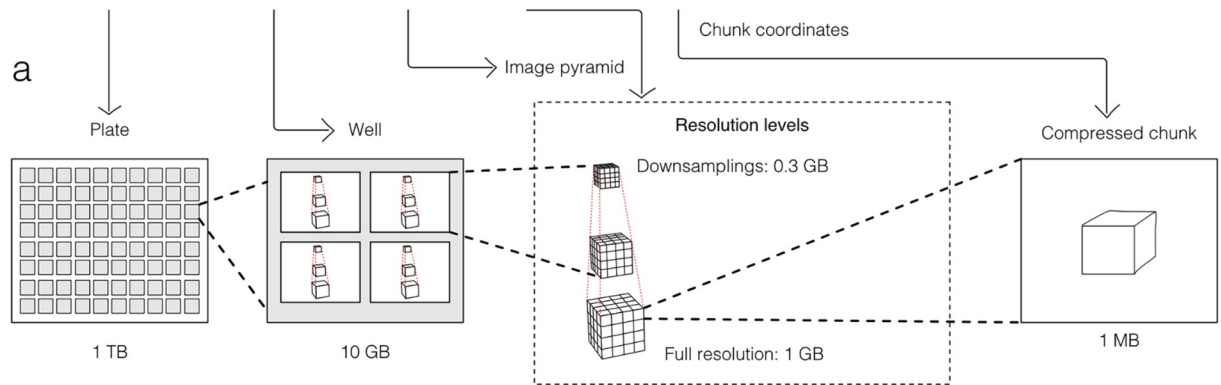
Raw images, classical features, deep-learning features



Chandrasekaran, S. N., Ackerman, J., Alix, E., Michael Ando, D., Arevalo, J., Bennion, M., Boisseau, N., Borowa, A., Boyd, J. D., Brino, L., Byrne, P. J., Ceulemans, H., Ch'ng, C., Cimini, B. A., Clevert, D.-A., Deflaux, N., Doench, J. G., Dorval, T., Doyonnas, R., ... Carpenter, A. E. (2023). JUMP Cell Painting dataset: morphological impact of 136,000 chemical and genetic perturbations. In *bioRxiv* (p. 2023.03.23.534023). <https://doi.org/10.1101/2023.03.23.534023>

NGFF: OME-Zarr

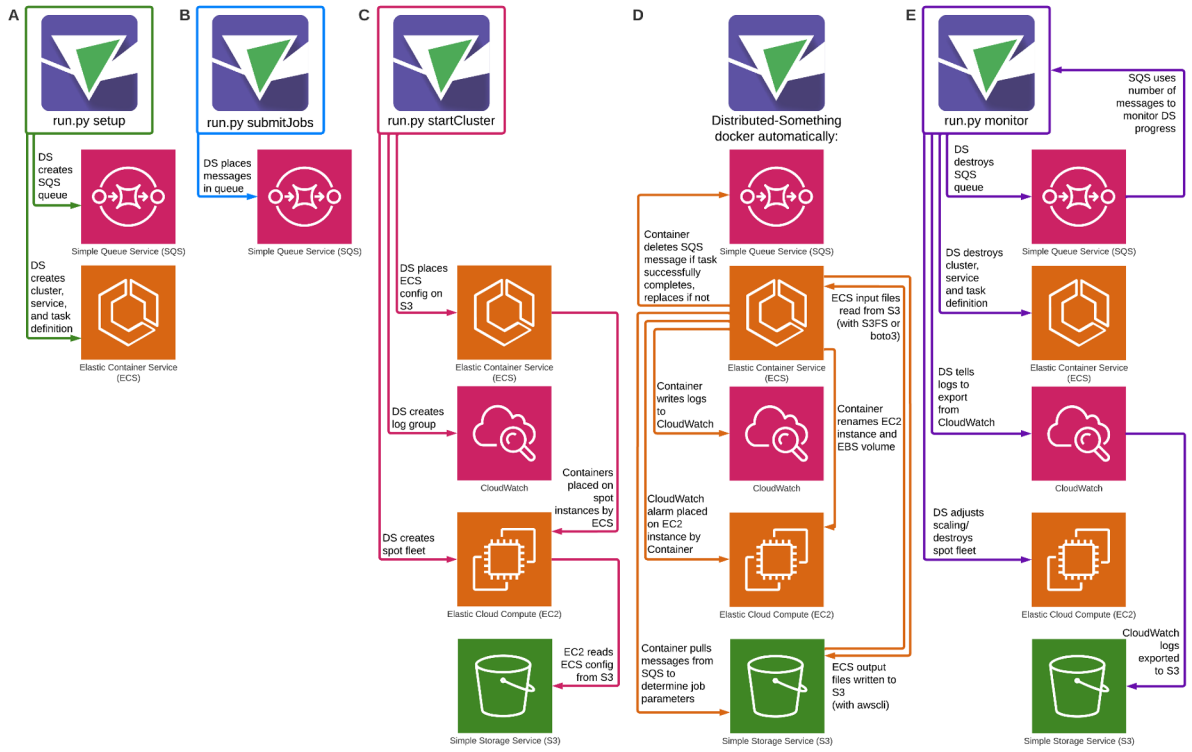
plate.ome.zarr/row/col/well/image/resolution/t/c/z/y/x




Moore, J., Basurto-Lozada, D., Besson, S., Bogovic, J., Bragantini, J., Brown, E. M., Burel, J.-M., Casas Moreno, X., de Medeiros, G., Diel, E. E., Gault, D., Ghosh, S. S., Gold, I., Halchenko, Y. O., Hartley, M., Horsfall, D., Keller, M. S., Kittisopikul, M., Kovacs, G., ... Swedlow, J. R. (2023). OME-Zarr: a cloud-optimized bioimaging file format with international community support. *Histochemistry and Cell Biology*. <https://doi.org/10.1007/s00418-023-02209-1>

Distributed-Something


Unlimited compute power/storage, low computational expertise



Weisbart, E., & Cimini, B. A. (2023). Distributed-Something: scripts to leverage AWS storage and computing for distributed workflows at scale. *Nature Methods*. <https://doi.org/10.1038/s41592-023-01918-8>

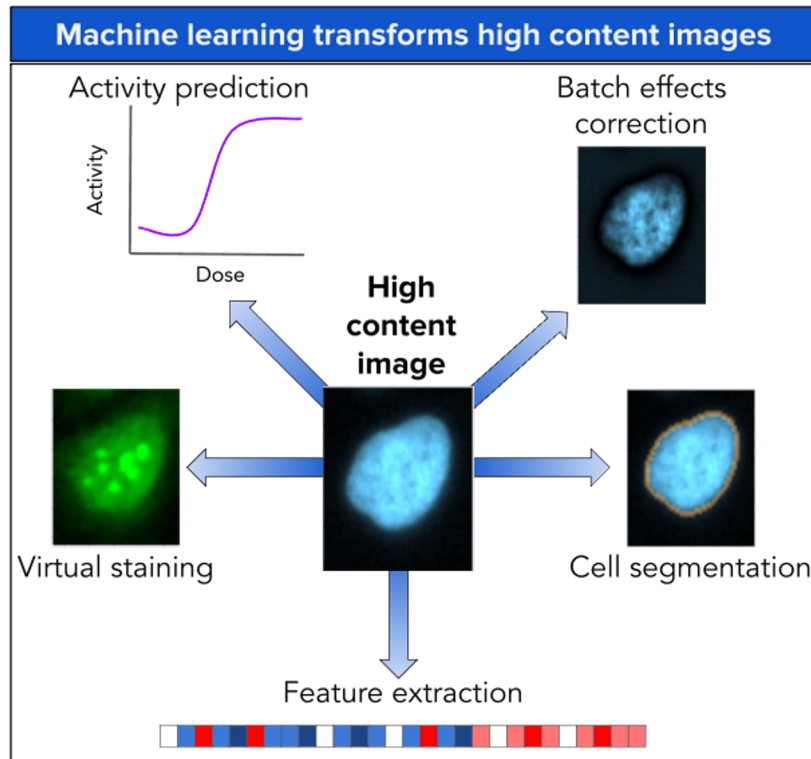



How has deep
learning changed
imaging?




Deep learning can help us get data from bioimages in many different ways

Caicedo and Cimini, arxiv 2023

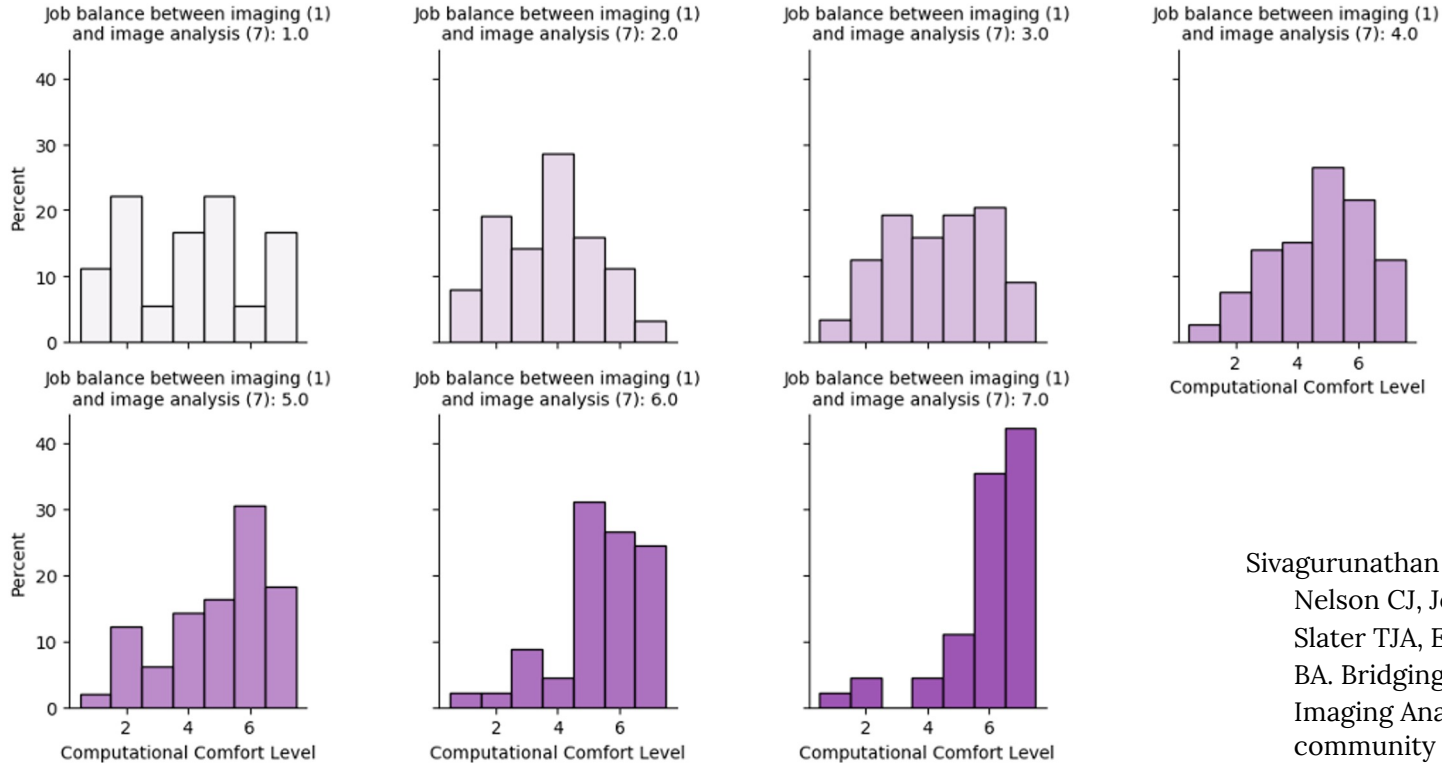




How can we bring
everyone into the
deep learning
revolution?

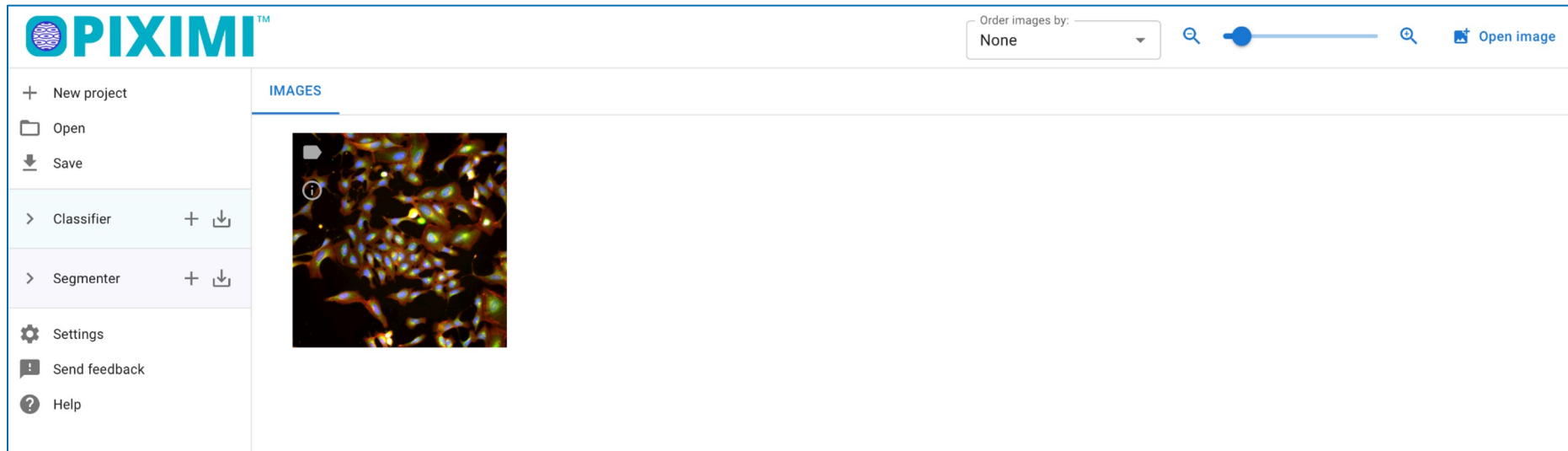


Scientists generating data are less computationally comfortable



Sivagurunathan S, Marcotti S,
Nelson CJ, Jones ML, Barry DJ,
Slater TJA, Eliceiri KW, Cimini
BA. Bridging Imaging Users to
Imaging Analysis - A
community survey. *J Microsc.*
2023 Sep 20.

PIXIMI: Images to Discovery



<https://www.piximi.app/>

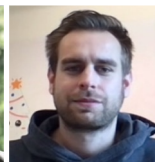
<https://github.com/piximi/piximi>



Allen
Goodman



Alice
Lucas



Levin
Moser



Nodar
Gogoberidze

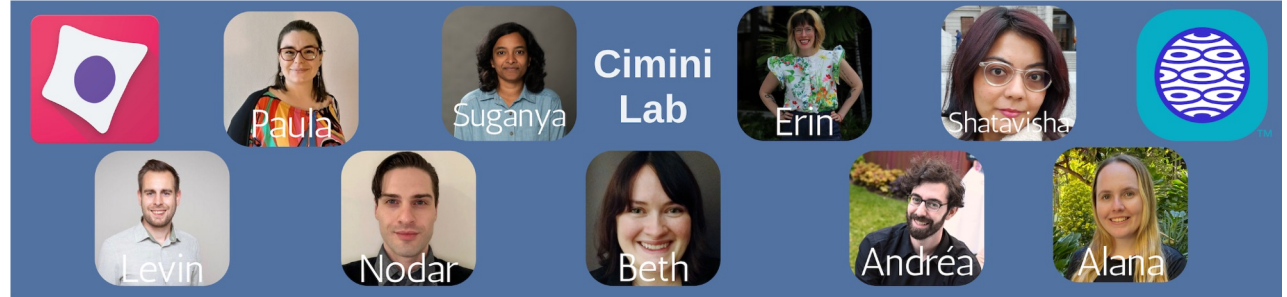


Andréa
Papaleo

Acknowledgements

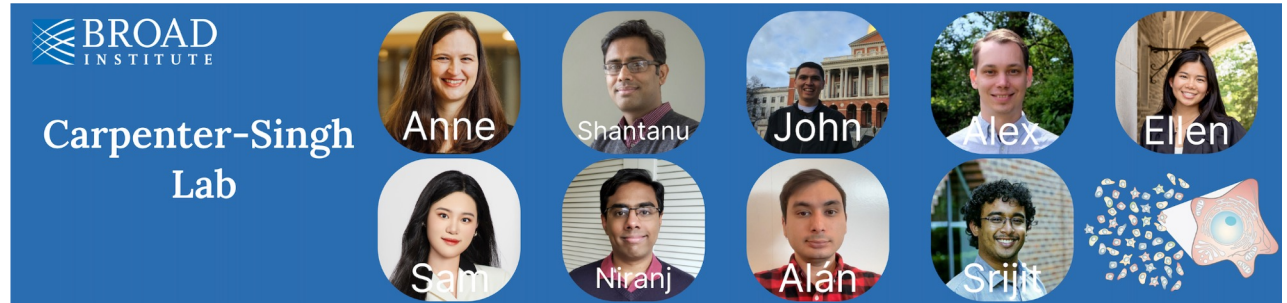
Cimini lab members

Beth Cimini
Shatavisha Dasgupta
Nodar Gogoberidze
Alana MacMahon
Levin Moser
Eddy Onyango
Andréa Papaleo
Suganya Sivagurunathan
Erin Weisbart



Carpenter-Singh lab members

Anne Carpenter
Shantanu Singh
John Arevalo
Niranj Chandrasekaran
Sam Chen
Alexander Kalinin
Ank Kumar
Alana MacMahon
Alán Muñoz González
Srijit Seal
Ellen Su



Recent major funding for this work provided by:

- CZI Imaging Scientist Fellowship
- NIH NIGMS: MIRA R35 GM122547
- CZI Software Fellows program
- NIH NIGMS: P41 GM135019
- Massachusetts Life Sciences Center